## AMENDMENTS TO THE SPECIFICATION

Please replace paragraph [0176] with the following paragraph:

**[0176]** Cloning of B7H3 full-length complementary DNA. A human ovarian cancer cell SKOV3 cDNA library constructed in Gateway™ pCMV•SPORT6 vector (Invitrogen, Carlsbad, CA) was transfected into Chinese Hamster Ovary (CHO) cells using standard protocol. Transfectants expressing BLA8 antigen on the cell surface were enriched by "panning" two times on BLA8 antibody. After enrichment, cells were lysed to release DNA. The crude lysate was used as template for the cloning of B7H3 by polymerase chain reaction (PCR). Briefly, a pair of B7H3-specific primers (sense: 5′-AGCCGCCTCACAGGAAGATGCT-3′ (SEQ 10 NO:1), and antisense : 5′-CCCTGGTCCTCATGGTCAGGCTAT-3′ (SEQ ID NO:2)) were designed to flank the open reading frame (ORF) of B7H3 based on the EST sequences available in the GeneBank. PCR was performed by combining 1.5 μl cell lysate with 50 pmol each of B7H3-specific primers, nuclease-free $H_2O$, 1X PCR buffer, 200 μM each of dNTPs, and 2.5 *U* Platinum High-Fidelity *Taq* DNA polymerase (Invitrogen, Carlsbad, CA). The reaction mixture was incubated at 94°C for 2 min, followed by 28 cycles of template denaturation at 94°C for 10 sec and primer annealing/extension at 68°C for 3 min, and a final extension at 68°C for 10 min. An aliquot of the PCR product was resolved onto a 1.2% agarose gel and visualized by ethidium bromide staining. A predominant band at about 1.6 kb was identified, gel-purified and cloned into pTargeT™ mammalian expression vector (Promega, Madison, WI). The transformants were characterized by restriction fragment analysis using *Hind*III, and the orientation of cDNA insert was determined by PCR. Clones with similar restriction patterns and correct orientation were selected for nucleotide sequencing. A 1605 bp ORF was discovered from seven of these clones, and all of them overlapped with B7H3 EST sequences available in the GeneBank. The authenticity of these cDNA clones were further confirmed by expressing them in CHO cells followed by staining with BLA8 antibody.

Please replace Table 2 on page 66 with the following:

**Table 2. Primers used in the detection of B7H3 mRNA transcript by RT-PCR**

| **Transfectant** | Primers used | **Approximate amplicon size** |
|---|---|---|
| Full-length B7H3 | sense : 5′-AGCCGCCTCACAGGAAGATGCT-3′ (SEQ ID NO:1)<br>antisense : 5′-AGCGGCCACCTGCAGGCTGACGGCA-3′ (SEQ ID NO:3) | 440 bp & 1.1 kb |
| ΔC | sense : 5′-AGCCGCCTCACAGGAAGATGCT-3′ (SEQ ID NO:1)<br>antisense : 5′-GGGGAATGTCATAGGCTGCCCTGTGAT-3′ (SEQ ID NO:4) | 760 bp |
| C | sense : 5′-CCCTACTCGAAGCCCAGCATGACCCT-3′ (SEQ ID NO:5)<br>antisense : 5′-CACGGCTCCTGTGGGGCTTCTCT-3′ (SEQ ID NO:6) | 330 bp |
| Δ5′ | sense : 5′-CCAGAGGCCCTGTGGGTGACCGT-3′ (SEQ ID NO:7)<br>antisense : 5′- CCCTGGTCCTCATGGTCAGGCTAT-3′ (SEQ ID NO:2) | 220 bp |
| Δ5′s | sense : 5′-CCAGAGGCCCTGTGGGTGACCGT-3′ (SEQ ID NO:7)<br>antisense : 5′- CCCTGGTCCTCATGGTCAGGCTAT-3′ (SEQ ID NO:2) | 220 bp |
| Δ3′ | sense : 5′- AGCCGCCTCACAGGAAGATGCT-3′ (SEQ ID NO:1)<br>antisense : 5′-CAGGGCTCCTGTGAGGCAGAACCA-3′ (SEQ ID NO:8) | 110 bp |
| Δ3′s | sense : 5′- AGCCGCCTCACAGGAAGATGCT-3′ (SEQ ID NO:1)<br>antisense : 5′-CACGGCTCCTGTGGGGCTTCTCT-3′ (SEQ ID NO:6) | 770 bp |